

Syllabus for B.Sc.(Biotechnology) SEP-2024

Semester I								
Sl. No	Course Code	Title of the Course	Category of Courses	Teaching Hours per Week	SEE	IA	Total Marks	Credits
1		Cell Biology and Genetics	Theory	4	80	20	100	3
2		Cell Biology and Genetics practical	Practical	4	40	10	50	2
Semester II								
3		Biochemistry and Biophysics	Theory	4	80	20	100	3
4		Biochemistry and Biophysics Practical	Practical	4	40	10	50	2
SEMESTER III								
5		Microbiology and Immunology	Theory	4	80	20	100	3
6		Microbiology and Immunology practical	Practical	4	40	10	50	2
SEMESTER IV								
7		Molecular Biology and Recombinant DNA Technology	Theory	4	80	20	100	3
8		Molecular Biology and Recombinant DNA Technology Practical	Practical	4	40	10	50	2
ELECTIVE COURSES - III								
9		Biotechnology in Daily life	Theory	2	40	10	50	2
ELECTIVE COURSES - IV								
10		Applications of Biotechnology in Agriculture	Theory	2	40	10	50	2

Core Courses

SEMESTER-I

Code :CELL BIOLOGY AND GENETICS

56 hours

Course Outcomes: After successful completion of this Course, students will be able to:

CO 1. Understand concepts of Biotechnology and demonstrate knowledge acquired in interdisciplinary skills in cell biology, genetics, biochemistry, microbiology, and molecular biology

CO 2. Describe the ultrastructure of cells, structure and function of organelles, cytosol and cytoskeleton

CO 3. Understand phases of cell cycle, cell division, reductional division in gametes, molecular mechanisms that regulate life and death of a cell including programmed cell death or apoptosis and cell cycle regulation.

CO 4. Comprehend organization and structure of chromosomes, and Mendelian laws of inheritance, deviations and exceptions to these laws.

CO 5. Describe mutations at the molecular level, types of mutations, genetic or hereditary disorders and concepts in population genetics

UnitI

(14 hours)

Discovery of cell, the cell Theory, classification of cell types. Levels of organization in cell biology: cell, tissue, organs and organism. Ultra structure of a eukaryotic cell- (Both plant and animal cells), Surface Architecture: Structural organization and functions of plasma membrane and cell wall of eukaryotes. Cellular Organelles: Structure and functions of cell organelles – Endoplasmic reticulum, Golgi complex, Mitochondria, Chloroplast, Ribosomes, Lysosomes, Peroxisomes, Nucleus (Nuclear envelope with nuclear pore complex, Nucleolus, Nucleoplasm and Chromatin). Vacuole, Cytosol and Cytoskeleton structures (Microtubules, Microfilaments and Intermediate filaments)

UnitII

(14 hours)

Chromosomes and cell division: General Introduction, Discovery, Morphology and structural organization –chromatids, Centromere, Secondary constriction, Telomere, Chromonema, Euchromatin and Heterochromatin, Chemical composition and Karyotype. Nucleosome organisation. Special type of chromosomes: Salivary gland and Lamp brush chromosomes.

Cell Division: Cell cycle, phases cell division. Mitosis and meiosis, regulation of cell cycles cell cycle checkpoints. Significance of cell cycle, synaptonemal complex. Cell Senescence and programmed cell death.

UnitIII

(14 hours)

Mendelian laws of inheritance - dominance, segregation, incomplete dominance, codominance with an example. Law of independent assortment, test cross, back cross. Deviations to Mendelian inheritance, complementary, supplementary and interaction of genes (13:3 ratio), epistasis, polygenic inheritance and multiple alleles.

Maternal Inheritance: Extrachromosomal inheritance, mitochondria and chloroplast genetic systems. Sex-linked inheritance, Chromosome theory of inheritance.

Sex determination in plants and animals: Concept of allosomes and autosomes, XX-XY, XX-XO, ZW-ZZ, ZO-ZZ types.

Unit IV

(14 hours)

Fine structure of gene-recon, muton and cistron.

Mutations: Types of mutations, Spontaneous and induced mutations, Mutagens chemical and physical mutagens, induced mutations in plants and microbes and its applications.

Structural and numerical aberrations of chromosomes; evolutions of wheat, and cotton

Hereditary defects: Klinefelter, Turner, Cri-du-Chat and Down syndromes. Analysis of mutations in Biochemical pathways, one gene - one enzyme hypothesis. Population genetics: Hardy-Weinberg equilibrium, gene, genotypes and gene frequencies.

References

Dale JW. 1990. *Molecular genetics of Bacteria*. John Wiley and Sons.

De Robertis EDP and De Robertis EMF. 1995. *Cell and Molecular Biology*. 8th edition, BI Waverly Pvt. Ltd., New Delhi.

Gardner et al. 2003. *Principles of Genetics*-8th edition. John Wiley and Sons, New York.

Gupta ML and ML Jangir. 2002. *Cell Biology-Fundamentals and Applications*. Argosies, Jodhpur, India.

Lewin B. 1994. *Genes VII* 5th edition. Oxford University Press, London.

Powar CB. *Cell Biology* 3rd edition. Himalaya Publishing House, Mumbai.

Rajeshwari S Setty and V. Sreekrishna. 2002. *Biotechnology-2* (Cell biology, Genetics, Microbiology). New Age International Publishers, New Delhi.

Taylor DJ, Green NPO and Stout GW. 1998. *Biological Science* 3rd Edition, Cambridge edition, Cambridge University Press, UK.

Sub Code**:CELL BIOLOGY AND GENETICS PRACTICAL**

1. Study and maintenance of simple and compound microscope
2. Use of Micrometer and calibration, measurement of onion epidermal cells
3. Study of divisional stages in mitosis from onion root tips
4. Study of divisional stages in meiosis in grasshopper testes/onion or Rheo flower buds.
5. Salivary gland chromosome isolation and staining.
6. Buccal smear – Barr bodies
7. Counting of RBC/WBC by Haemocytometer.
8. Karyotype analysis
9. Isolation of chloroplast.
10. Study of drosophila.
11. Separation of drosophila eye pigment by paper chromatography
12. Simple genetic problems based on theory

SEMESTER-II

Sub Code :BIOCHEMISTRY AND BIOPHYSICS

56 hours

Course Outcomes (COs): At the end of the course the student should be able to:

CO 1. Acquire knowledge about types of biomolecules, structure, and their functions

CO 2. Will be able to demonstrate the skills to perform bioanalytical techniques

CO 3. Apply comprehensive innovations and skills of biomolecules to biotechnology field

Unit I

(14 hours)

Aims and scope of biochemistry and biophysics

General classification, structure and functions of biomolecules: carbohydrates, proteins, lipids and nucleic acids. Detailed structure and general properties of monosaccharides, glycosidic bond; structural polysaccharides - cellulose, chitin, peptidoglycans; storage polysaccharides - starch, glycogen. Classification of standard amino acids, peptide bonds, general properties of amino acids, titration curve. Protein structure - primary, secondary, tertiary and quaternary with examples.

Unit II

(14 hours)

Enzymes: History, general properties, active site, Michaelis-Menten equation, allosteric enzymes; nomenclature and classification. Enzyme inhibition types- reversible, non-competitive and uncompetitive with examples. Multienzyme and isoenzyme with examples. Brief account of applications of enzymes: enzymes in genetic engineering - restriction enzymes and polymerases; enzymes in clinical significance - LDH, SGOT, SGPT and diagnostic kits.

Unit III

(14 hours)

Structure and function of water, pH impact on biomolecular reactions, Henderson and Hasselbach's equation with applications. Buffers- types and applications. Laws of thermodynamics, free energy, ATP as biological energy currency.

Lambert - Beer's law, absorption spectrum, absorption maxima. UV-Vis spectroscopy; mass spectroscopy, atomic absorption spectroscopy with applications. Brief account of principles and applications to understand the structure of molecules: X-ray crystallography and NMR.

Unit IV

(14 hours)

Microscopy: Magnification, Resolution power, Optical - Bright field, dark field, phase contrast and fluorescence; Electron microscopy - TEM and SEM. Partition coefficient, Paper and thin layer chromatography-principle, methodology, applications and significance of R_f value. Gel filtration chromatography, affinity chromatography, gas liquid chromatography and high performance liquid chromatography. Principle, procedure and applications of electrophoresis- agarose and polyacrylamide gel electrophoresis. Centrifugation: differential, density gradient and ultra - principle, instrumentation and applications

References

- AroraMP.2007.*Biophysics*.Himalaya PublishingHouse,NewDelhi.
- Bialek W. 2012. *Biophysics: Searching for Principles*. Princeton University Press.
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- LeningerAL,NelsonDLandCoxMM.1993.*PrincipleofBiochemistry*.CBSPublications. Nickolas CPL.1982. *Fundamentals of Enzymology*. Oxford Publishers.
- StryerL.1988.*Biochemistry*3rdedition,Freeman&Co,New York.
- Trevor.Enzymebiochemistry,BiotechnologyandClinicalChemistry.HarwoodPublishers.
- UpadhyayandUpadhyay A.2000.*BiophysicalChemistry-PrinciplesandTechniques*. HimalayaPublishers,NewDelhi,
- ZubayJ.1988. *Biochemistry*2ndedition.MacMillanPublishingCompany,NewYork.

Sub Code :BIOCHEMISTRY AND BIOPHYSICS PRACTICAL

1. Introduction to basic instruments (Principle, standard operating procedure) with demonstration.
2. Microscopy
3. Preparation of standard buffers– Citrate and phosphate, determination of pH of solution using pH meter.
4. Qualitative tests for carbohydrates-monosaccharides, disaccharides and polysaccharides.
5. Qualitative tests for proteins.
6. Estimation of maltose by DNS method
7. Determination of α -amylase activity by DNS method
8. Estimation of proteins by Lowry's and Biuret method
9. Lambert – Beer's law
10. Absorption maxima of a solution
11. Paper chromatography
12. Thin layer chromatography
13. Gel Electrophoresis

SEMESTER-III

Sub Code :MICROBIOLOGY AND IMMUNOLOGY

56 hours

After successful completion of this Course, students will be able to:

- CO 1. Understand Historical prospective and scope Microbiology
- CO 2. Experiment with various methods of sterilization in microbiological work
- CO3. Prepare different types of media, perform culture methods, preservation of microorganisms for isolation, characterization of microbes
- CO 4. Handle and use antimicrobial agents and perform anti-microbial assays
- CO 5. Demonstrate the Laboratory skills in basic and applied microbiology with reference to technological aspects.
- CO6. Demonstrate comprehension of the underlying structure and function of the immunesystem and related disorders.
- CO 7. Demonstrate an understanding of the role of cells and molecules in immune reactions and responses
- CO8. Demonstrate technical skills in immunological tools and techniques
- CO 9. Explain the fundamental concepts of immunity, and the contributions of the organs and cells in immune responses.
- CO 10. Realize how the MHC molecule's function and host encounters an immune insult.
- CO 11. Understand the antibodies and complement system
- CO12. Comprehend the overreaction by our immune system leading to hypersensitive conditions and its consequences

UnitI (14 hours)

Aim, scope and historical perspectives of microbiology. Contributions of early microbiologists: Leeuwenhoek, Louis Pasteur, Robert Koch and Edward Jenner. General classification of microbial kingdom - classical, nutritional and molecular approaches. Concepts of sterilization and disinfection: dry heat, moist heat, radiation, chemical and filtration. Introductory concepts in virology- classification, structure and life cycle - lysogenic and lytic cycle. Antibiotics: classification and mode of action.

UnitII (14 hours)

Prokaryotic and eukaryotic microbes and their characteristics. Structure and functions of microbial cell wall, lipopolysaccharides, flagella, capsules, endospores, pili (fimbriae), cell membranes and cell inclusions.

Culture Media: Components of media, natural and synthetic media, selective, differential, indicator, enriched media.

Pure culture methods: Serial dilution and plating methods (pour, spread, streak); maintenance and preservation/stocking of pure cultures

Microbes in extreme environments- thermophiles, psychrophiles, acidophiles, alkaliphiles, halophiles and barophiles. Microbe -microbe interactions.

UnitIII (14 hours)

Historical perspectives in immunology, Origin and diversity of immunology. Classification of immunity: innate and adaptive immunity. Immune systems: organs, cells of immune system. Role of B cell and T cell in humoral and cell mediated immunity. Major histocompatibility complexes (MHCs)-types, structure and their functions. Structure, types and functions of antigens. Haptens and adjuvants, Structure, types and functions of antibodies.

UnitIV (14 hours)

Antigen-antibody reactions - precipitin test, agglutination test, complement fixation (or complement cascade) reaction and ELISA. Immunoblot - types, principle and applications. Immunological disorders - autoimmune diseases - Brief descriptions of two autoimmune diseases- Rheumatoid arthritis and myasthenia gravis. Acquired immunodeficiency syndrome: description of causative agent, and briefly description of mechanisms. Hypersensitivity and allergy. Vaccines-classical and modern.

References

- Abbas A, Lichtman A and Pillai S. 2015. *Cellular and Molecular Immunology*. Elsevier Saunders Co.
- Brock TB and Madigan. 1988. *Biology of Microorganisms*. Prentice Hall, New Jersey.
- Cassida, L.E. 1968. *Industrial Microbiology* John Wiley & Sons.
- Ivan Riott Jonathan Brostoff and David Male. *Immunology* 3rd edition. Mosby Publishers.
- Janeway and Travers. *Immunobiology* 3rd edition. Churchill Livingstone Publications.
- Kuby J. 2000. *Immunology*. Freeman and Company Publishers.
- Maloy SR. Friefelder 1994. *Microbial Genetics*. Jones and Bartlett Publishers.
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- Subba Rao N.S. 1974. *Soil Microbiology*, 4th edition, Oxford & IBH Publishers, New Delhi
- Torture GJ, Frank BR, and Case CL. 1992. *Microbiology- An Introduction*. Communing Publishing Company Inc, California.

Sub Code :MICROBIOLOGY AND IMMUNOLOGY PRACTICAL

1. Instrumentation and microscopy with special reference to microbiology and immunology.
2. Sterilization techniques – dry heat sterilization with hot air oven, wet heat sterilization with autoclave, membrane filtration and assessment for sterility
3. Preparation of culture media for bacteria, fungi and their cultivation
4. Plating technique – pour plate and spread plate
5. Isolation of microorganisms - air, water, human body and soil.
6. Study of bacterial colony characteristics.
7. Purification of bacterial and fungal cultures using streak plate technique/mycelial transfer
8. Culture preservation techniques – slant and stab culture
9. Staining of microorganisms - Gram staining, capsule staining, spore staining, negative staining.
10. Biochemical activities of microorganisms - indole, methyl red, Voges-Proskauer and catalase tests. Starch hydrolysis, Catalase test, Gelatin hydrolysis
11. Antibiotic sensitivity of microorganisms.
12. Blood grouping and Rh factor determination.
13. Study of different types of leucocytes.
14. Immunodiffusion studies.
15. Dot ELISA

SEMESTER-IV

Sub Code : MOLECULAR BIOLOGY AND RECOMBINANT DNA TECHNOLOGY

56 hours

Course Outcomes (COs): At the end of the course the student should be able to:

CO 1. Study the advancements in molecular biology with latest trends.

CO 2. Will acquire the knowledge of structure, functional relationship of proteins and nucleic acids.

CO 3. Aware about the basic cellular processes such as transcription, translation, DNA replication and repair mechanisms.

CO 4. Demonstrate a thorough understanding of the fundamental principles and techniques of genetic engineering.

CO 5. Apply the knowledge of genetic engineering to diverse applications in agriculture, medicine, biotechnology, and environmental science.

CO 6. Perform laboratory procedures and develop practical skills in genetic engineering techniques.

CO 7. Evaluate genetic engineering's ethical, social, and legal implications and propose responsible solutions.

CO 8. Stay updated with recent advancements in genetic engineering, critically evaluate emerging trends, and assess their potential impact on various fields.

Unit I

(14 hours)

Discovery, structure and types of DNA and RNA. Experiments on DNA as genetic material. Replication of DNA in prokaryotes and eukaryotes. – Enzymes and proteins involved in replication, Theta model and rolling circle model. Polymerases and all enzyme components. Mechanisms of DNA recombination in prokaryotes and eukaryotes. DNA damage and Repair mechanism: photo reactivation, excision repair, mismatch repair and SOS repair.

Unit II

(14 hours)

Structure of prokaryotic and eukaryotic genes. Central dogma of molecular biology. Transcription in prokaryotes RNA polymerase, role of sigma factor, promoter, Initiation, elongation and termination of RNA chains. Transcription in eukaryotes: Eukaryotic RNA polymerases, mechanism of transcription initiation, promoter clearance, elongation and termination. Post transcriptional modification. RNA splicing and processing: Genetic code and its characteristics, Translation- in prokaryotes and eukaryotes. Prokaryotic gene expression- *lac* and *trp*.

Unit III

(14 hours)

Aims, objectives and scope of gene cloning and steps involved recombinant DNA technology. Isolation and purification of DNA from bacterial, plant and animal cells. Tools of DNA modification: restriction enzymes- properties, classification, types with examples; ligation, DNA modifying enzymes. DNA vectors: plasmids, bacteriophages, cosmids, BAC, YAC, plant and animal viruses. Gene transfer technique.

Unit IV

(14 hours)

Genomic and cDNA libraries: features, construction and application. Screening and selection of recombinants by selection media, insertional inactivation. PCR- principle, protocol and applications. DNA sequencing- Sanger's, next generation sequencing. Gene editing techniques principle and applications. Blotting techniques: Southern, Northern and Western- principle and applications. Probes - types, preparation and application. DNA finger printing- principle and applications. Hazards and biosafety measures for recombinant DNA technology and GMOs.

References

- Alberts B, Bray D, Lewis J, Raff M, Roberts K and Watson JD. 2002. *Molecular Biology of the Cell* 4th edition. Garland Publishing, Inc., New York.
- Cooper GM. 2000. *The Cell - A Molecular Approach* 2nd edition. Sunderland (MA): Sinauer Associates, Inc.
- De Robertis EDP and De Robertis EMF. 1995. *Cell and Molecular Biology*. 8th edition, BIWaverly Pvt.Ltd., New Delhi.
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Sub Code

**:MOLECULAR BIOLOGY AND RECOMBINANT DNA
TECHNOLOGY PRACTICAL**

1. Preparation of DNA model
2. Isolation of DNA from bacteria, plant and animal tissues.
3. Isolation of RNA
4. Tests for DNA/RNA/proteins isolated from tissues
5. Estimation of DNA by DPA method
6. Estimation of RNA by Orcinol method
7. Column chromatography – gel filtration (Demo)
8. Agarose gel electrophoresis to separate DNA
9. SDS PAGE
10. ligation
11. Restriction digestion
12. Bacterial transformation
13. Western blotting technique

Syllabus for B.Sc.(Biotechnology)
CBCS-2025
ELECTIVE COURSES

SEMESTER III

BSCBTOE283: BIOTECHNOLOGY IN DAILY LIFE

28 hours

Unit I

(14 hours)

Definition and History of Biotechnology, Scope and Importance of Biotechnology. Microbial world, fermentation. Applications of Biotechnology in Industry: Production of citric acid, alcoholic beverages, Enzymes like proteases, lipases and amylases. Plant biotechnology- GM crops, gene transfer technology, bioreactors, disease control through Bt genes. Applications of Biotechnology in Agriculture: Biofertilizers, Biopesticides, Transgenic plants, Mushroom production

Unit II

(14 hours)

Applications of Biotechnology in Medicine and pharmaceuticals: Insulin therapeutic molecules like tPA, factor VIII, antibiotics. Animal biotechnology – transgenic animals, test tube babies (*In-vitro* fertilization), ethical issues, animal bioreactors, stem cells, stem cell therapy, Environmental biotechnology- composting, biodegradation, biotransformation, biomining. Applications of Biotechnology in Environmental pollution control: Municipal Solid waste management, sewage and industrial effluent treatment, biofuels, Petroleum degradation.

References

- Brock TB and Madigan. 1988. *Biology of Microorganisms*. Prentice Hall, New Jersey.
- Bajaj YPS. 2007. *Biotechnology in Agriculture and Forestry*. Springer Verlag Publishers.
- Chawla HS. 2004. *Introduction to Plant Biotechnology*. Science Publications. Inc.
- Houdebine LM. 2003. *Animal Transgenesis and Cloning*. John Wiley & Sons.
- Ian Freshney R. 2000. *Culture of Animal Cells: A Manual of Basic Technique* 4th edition. Wiley-Liss.
- Jay JM, Loessner MJ and Golden DA. 2005. *Modern Food Microbiology*. Springer Publication, New York.
- Jogdand SN. 2010. *Environmental Biotechnology*. Himalaya Publishing House, Bangalore, New Delhi.
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- Prave P. et al. 1987. *Fundamentals of Biotechnology*. Wiley-Black well Publications.
- Russell, G.E. 1988. *Biotechnology of Higher Plants*. Intercept Publications.
- Subba Rao N.S. 1974. *Soil Microbiology*, 4th edition, Oxford & IBH Publishers, New Delhi

SEMESTER IV

BTC 302 APPLICATIONS OF BIOTECHNOLOGY IN AGRICULTURE

28 hours

Course Outcomes:

After successful completion of this Course, students will be able to:

CO1. Understand the biotechnological applications in agriculture

CO2. Understand the importance of biotechnological methods such as plant tissue culture

CO 3. Comprehend the pros and cons of GM crops and their plant products

CO4. Appreciate the biotechnological applications for effective pest control and crop improvements

Unit I

(14 hours)

Agricultural Biotechnology: Concepts and scope of biotechnology in Agriculture. Plant tissue culture, micro propagation, entrepreneurship in commercial plant tissue culture. Banana tissue culture—primary and secondary commercial setups, Small scale bioenterprises: Mushroom cultivation.

Unit II

(14 hours)

Transgenic plants: The GM crop debate—safety, ethics, perception and acceptance of GM crops GM crops case study: Bt cotton, Bt brinjal. Plants as biofactories for molecular pharming; edible vaccines, plantibodies, nutraceuticals. Genetic engineering for quality improvement: Seed storage proteins, Flavours- capsaicin, vanillin.

References:

Chrispeels M.J. and Sadava D.E.(1994) Plants, Genes and Crop Biotechnology, 2nd Ed., Jones and Bartlett Publishers, Boston.

Gamborg O.L. and Philips G.C.(1998) Plant cell, tissue and organ culture, 2nd Ed., Narosa Publishing House. New Delhi.

Gistou, P. and Klu, H.(2004). Hand book of Plant Biotechnology (Vol.I&II). John Publication.

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Kyte, L., Kleyn, J., Scoggins, H., and Bridgen M.(2003) Plants from test tubes. An introduction to micropropagation, 4th Ed., Timber Press, Portland.

Murray D.R. (1996) Advanced methods in plant breeding and biotechnology. Panima Publishing Corporation.

Nickoloff, J.A. (1995). Methods in molecular biology, Plant cell electroporation and electrofusion protocols-Humana Press Incorp, USA.

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